

INTRACELLULAR UPTAKE AND TOXICITY OF GOLD NANOPARTICLES (AuNPs) IN REPRESENTATIVE CELL LINES

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19 November 2020







BACKGROUND What are NPs?

- Nanoparticles (NPs) or Engineered nanomaterials (ENM) are microscopic entities having at least one dimension 1-100 nm in size.
- The most commonly used nanoparticles (NPs) are titanium dioxide (TiO₂), zinc oxide (ZnO), silver (Ag), carbon nanotubes (CNTs), silicon dioxide (SiO₂) and gold (Au).







BACKGROUND

Characteristics of AuNPs



Physiochemical properties of AuNPs

NPs do not behave like their bulk counterparts
posses unique properties









BACKGROUND

Applications of AuNPs



Modified AuNPs can be used in bio- applications such as:





• Some NPs readily travel throughout the body, deposit in target organs, penetrate cell membranes and may prompt injurious responses.



BACKGROUND



- Common routes of exposure
 - inhalation
 - intravenous injection
 - ingestion
 - transdermal delivery
- Human skin keratinocyte (HaCaT) cells are used to investigate the effects of NPs on skin
- The boundary to AuNPs entering the body through direct ingestion is the gastrointestinal (GI) system
 - human epithelial colorectal adenocarcinoma (Caco-2) cells can been used as a general *in vitro* model for intestinal cells.
- Exposure of NPs to various cell lines may lead to uptake of these nanomaterials.







AIM and OBJECTIVES

Aim

• The purpose of this study was to investigate the effects of different functional groups, present on ligands bound to the surface of AuNPs, on their intracellular uptake, cytotoxicity and genotoxicity in the human epithelial colorectal adenocarcinoma cells (Caco-2) and the human skin keratinocyte cells (HaCaT) cells.

Objectives

- To assess the uptake of 14 nm citrate-stabilized (CITR14) AuNPs and 14 nm PEG-liganded amine (PEG-NH₂14) AuNPs using the CytoViva darkfield hyperspectral imaging (HSI) system.
- To assess the cytotoxicity of CITR14 AuNPs and PEG-NH₂14 AuNPs using the xCELLigence system.
- To assess the genotoxicity of CITR14 AuNPs and PEG-NH₂14 AuNPs using the *in vitro* micronucleus assay.



METHODOLOGY NP Uptake



Cell culture

- Caco-2 and HaCaT cells supplemented with 10% fetal bovine serum and 1% penicillin streptomycin
- The CytoViva hyperspectral imaging (HSI) system is a technique used for assessing NP intracellular distribution and it can also confirm the presence of NPs within the cells







METHODOLOGY









METHODOLOGY

Genotoxicity



Micronucleus (MN) assay

- MN assay quantifies subnuclear bodies- micronuclei
 - detects chromosome breakage and changes in chromosome number
- Treated cells were grown for 20 hrs to allow chromosome or spindle damage
 - leading to formation of micronuclei (MNi), nucleoplasmic bridges (NPBs) and nuclear buds (NBuds)





Fig. 2. Principles of the MN assay (Fenech et al., 2007)





AuNP uptake

AuNP uptake by Caco-2 cells



Fig. 3. Caco-2 cells treated with CITR14 AuNPs and PEG-NH₂14 AuNPs at concentrations of 1 nM (left panel) and 5 nM (right panel) and for the negative control, cells were not exposed to AuNPs. Cells were exposed to AuNPs for 24 hrs.

Fig. 4. HaCaT cells treated with CITR14 AuNPs and PEG-NH₂14 AuNPs at concentrations of 1 nM (left panel) and 5 nM (right panel) and for the negative control, cells were not exposed to AuNPs. Cells were exposed to AuNPs for 24 hrs.

AuNP uptake by HaCaT cells





Cytotoxicity of AuNPs

Cytotoxic effects of CITR14 AuNPs in Caco-2 cells



5 nM; 2 nM; 1 nM; 0.5 nM; Untreated

Fig. 5. The cytotoxicity of CITR14 AuNPs in the Caco-2 cells indicated by the normalized CI over a period of 100 hours. Error bars represents SD between the replicates.

 A similar CI for cells treated with 0.5 nM, 1 nM, 2 nM CITR14 AuNPs and the untreated cells, while 5 nM induced significant (p < 0.01) CI decrease

Cytotoxic effects of CITR14 AuNPs in HaCaT cells





Fig. 6. The cytotoxicity of CITR14 AuNPs in the HaCaT cells indicated by the normalized CI over a period of 100 hours. Error bars represents SD between the replicates.

- 5 nM reduced CI the most and showed a statistically significant difference (p < 0.001) from the untreated control.
- No significant difference (p > 0.05) between other treatments and the control.





Cytotoxicity of AuNPs

Cytotoxic effects of PEG-NH₂14 AuNPs in Caco-2 cells



5 nM; **2** nM; **1** nM; **0.5** nM; **Untreated**

Fig. 7. The cytotoxicity of PAM14G AuNPs in the Caco-2 cells indicated by the normalized CI over a period of 100 hours. Error bars represents SD between the replicates.

 No significant difference was observed between the cells treated with PEG-NH₂14 AuNPs and the untreated control (p > 0.05) Cytotoxic effects of PEG-NH₂14 AuNPs in HaCaT cells





Fig. 8. The cytotoxicity of PEG-NH₂14AuNPs in the HaCaT cells indicated by the normalized CI over a period of 100 hours. Error bars represents SD between the replicates.

 All concentrations increased CI, there was no statistical difference (p > 0.05) between the treatments and the untreated control.





RESULTS AND DISCUSSIONS Genotoxicity of AuNPs

- Different types of chromosome aberrations
- Visualized and scored using the white light microscope at 100 X magnification



Fig. 4.19. Different types of chromosome aberrations formed in the Caco-2 and HaCaT cells using the MN assay. (A) Untreated binucleated Caco-2 cells with no aberrations, (B) binucleated cells with a NPB, (C) binucleated cell with a MNi and (D) binucleated cell with a NBud





Genotoxicity of AuNPs



Genotoxic effects of CITR14 AuNPs in Caco-2 cells

Fig. 10. Genotoxicity of CITR14 AuNPs at various concentrations. Ethyl methanesulfonate (EMS) was used as a positive control. Error bars represents mean SD.

- 2 nM and 5 nM AuNPs induced statistically significant (p < 0.001) MNi
- NPBs 1 nM and 5 nM (p < 0.01) and 2 nM (p < 0.001)
- All treatments induced significant nuclear buds (p < 0.05).

Genotoxic effects of CITR14 AuNPs in HaCaT cells



Fig. 11. Genotoxicity of CITR14 AuNPs at various concentrations. EMS was used as a positive control. Error bars represents mean SD.

- MNi 1 nM (P < 0.01), 2 nM and 5 nM (P < 0.001)
- NPBs all treatments (p < 0.05)
- NBuds all treatments (p > 0.05)





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Genotoxicity of AuNPs





Fig. 12. Genotoxicity of $PEG-NH_214$ AuNPs at various concentrations. Ethyl methanesulfonate (EMS) was used as a positive control. Error bars represents mean SD.

- MNi 1 nM and 2 nM (p < 0.05) and 5 nM (p < 0.001)
- NPBs 2 nM and 5 nM (p < 0.01)
- Buds all treatments (p > 0.05)

Genotoxic effects of PEG-NH₂14 AuNPs in HaCaT cells



Fig. 13. Genotoxicity of PEG-NH $_2$ 14 AuNPs at various concentrations. EMS was used as a positive control. Error bars represents mean SD.

• MNi - 5 nM (p < 0.001)

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- NPBs- all treatments (p > 0.05).
- Buds 2 nM and 5 nM (p < 0.01)





CONCLUSIONS

- The citrate-stabilized AuNPs entered both the Caco-2 and HaCaT cell lines at a higher level compared to PEGylated AuNPs.
 - This suggests that PEGylation discourages AuNP uptake
- CITR14 AuNPs showed a significant reduction in CI at the highest concentration of 5 nM in Caco-2 and HaCaT cells, therefore indicating cytotoxicity.
- In contrast, PEG-NH₂14 AuNP induced cell proliferation indicating that the amine functional group possibly enhance cell proliferation.
- All types of chromosomal aberrations (MNi, NPBs and NBuds) were formed by both the AuNPs in both cell lines.

- NPBs abundant

- Therefore, non-cytotoxic concentrations have the ability to induce genotoxic effects in Caco-2 and HaCaT cell lines.
- Suggesting that DNA damage was due to indirect mechanisms, such as the production of reactive oxygen species (ROS).



FUTURE WORK



- Gene expression profiles of AuNP treated cells should be determined to show the regulation of genes and growth factors that affect cell proliferation.
- Effects of different chemical functional groups, upon cell entry and toxicity must be investigated.
- ROS generation must be investigated to ascertain the possibility of oxidative DNA damage being the cause of the genotoxicity seen in this study.







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